



Annexin V-FITC/PI Apoptosis Detection Kit

Cat #: KTA0006

Size: 20 T/50 T/100 T

	Annexin V-FITC/PI Apoptosis Detection Kit		
REF	Cat #: KTA0006	LOT	Lot #: Refer to product label
	Applications: Flow cytometry and fluorescence detection for cell samples		
	Fluorescence Excitation/ Emission: Annexin V-FITC: E _x /E _m =488/525 nm, PI: E _x /E _m =535/617 nm		
	Note: Stored at -20°C for 12 months, protected from light		

Assay Principle

Apoptosis is a form of programmed cell death to remove unwanted, damaged, or senescent cells from tissues. In normal cells, the negative phospholipids reside on the inner side of the cellular membrane while the outer surface of the membrane is occupied by uncharged phospholipids (PS). After a cell has entered apoptosis, the negatively charged PS are transported from the inner to the outer leaflet of the plasma membrane, thus exposing PS to the external cellular environment. The human anticoagulant, Annexin V, is a 35-36 kDa Ca²⁺ dependent phospholipid-binding protein that has a high affinity for PS. Annexin V labeled with a fluorophore or biotin can identify apoptotic cells by binding to PS exposed on the outer leaflet. Propidium iodide (PI) is a fluorescent nucleus dye, impermeant to live cells and apoptotic cells, but stains dead cells with red fluorescence, binding tightly to the nucleic acids in the cell. Annexin V-FITC/PI Apoptosis Detection kit provides a rapid and convenient assay for apoptosis. After staining a cell population with Annexin V-FITC/PI in the provided binding buffer, early apoptotic cells show green fluorescence of the cellular membrane, dead cells show red fluorescence of the nucleus and green fluorescence of the cellular membrane, and live cells show little or no fluorescence. In addition, to address the non-standardization of apoptosis detection and results, the kit also provides a patented positive reference substance for apoptosis. Detection can be analyzed by flow cytometry or by fluorescence microscopy.

Materials Supplied and Storage Conditions

Kit components	Size			storage condition
	20 T	50 T	100 T	
Annexin V Binding Buffer (5×)	2 mL	5 mL	10 mL	4°C
Annexin V-FITC	100 µL	250 µL	500 µL	-20°C, protected from light
Propidium Iodide (PI)	40 µL	100 µL	200 µL	-20°C, protected from light
Apoptosis Inducer A	5 µL	5 µL	10 µL	-20°C
Apoptosis Inducer B	5 µL	5 µL	10 µL	-20°C

Materials Required but Not Supplied

- Centrifuge, fluorescence microscopy or flow cytometer

- Precision pipettes, disposable pipette tips, deionized water, glass slides
- Cell culture plate

Reagent Preparation

Note: Briefly centrifuge small vials at low speed prior to opening.

1×Annexin V Binding Buffer: Prepared before use. Dilute Annexin V Binding Buffer (5×) to 1×Annexin V Binding Buffer with deionized water. Store the surplus reagent at 4°C for 6 months.

Annexin V-FITC: Ready to use as supplied. Equilibrate to room temperature before use. The unused reagents are sub-packaged and stored at -20°C, protected from light. Avoid repeated freezing and thawing.

Propidium Iodide (PI): Ready to use as supplied. Equilibrate to room temperature before use. The unused reagents are sub-packaged and stored at -20°C, protected from light. Avoid repeated freezing and thawing.

Apoptosis Inducer A: Ready to use as supplied. Equilibrate to room temperature before use. Store at -20°C.

Apoptosis Inducer B: Ready to use as supplied. Equilibrate to room temperature before use. Store at -20°C.

Assay Procedure

I Positive cell apoptosis induction

1. For cultured cells to be induced apoptosis, the Apoptosis Inducer A or Apoptosis Inducer B were added into the culture medium according to the volume ratio of 1:1,000-1:3,000 (Apoptosis Inducer A and Apoptosis Inducer B can be added into the culture medium together).

2. Cell apoptosis was observed after 4, 8, 12, 16 or 24 h. For the most part, after 16-24 h, obvious changes in cell morphology can be seen under the light microscope, which can be used to observe apoptosis staining (It is recommended to adjust the induction time and concentration for different cells).

II Detection cell apoptosis by Annexin V-FITC/PI

A. Quantification by Flow Cytometry

1. Induce apoptosis in cells using the desired method. Prepare a negative control by incubating cells in the absence of inducing agent.

2. Collect $1-2 \times 10^5$ cells by centrifugation (4°C, 300 g, 5 min) and wash with ice-cold PBS twice.

Note: For adherent cells, using trypsin (EDTA free) to digest cells firstly and then centrifugation. The time of trypsinization should not be too long, because trypsin could destroy the membrane structure.

3. Resuspend the cells in 500 μ L 1×Annexin V Binding Buffer.

4. Add 5 μ L Annexin V- FITC and 2 μ L PI to each 500 μ L of cell suspension and mix gently.

5. Incubate the cells at room temperature for 15 min in the dark.

6. After the incubation period, keep the samples on ice. Analyze the cells by flow cytometry within 30 min of staining. Use 488 nm and 535 nm excitation and measure fluorescence emission near 525 nm (FITC channel) and 617 nm (PE or PI channel).

B. Detection by Fluorescence Microscopy

1. For suspension cells

1) Follow the protocol for flow cytometry from step A.1 to step A.6.

2) Place the cell suspension from Step A.6 on a glass slide. Cover the cells with a glass coverslip. Analyze cells by fluorescence microscopy using the appropriate filters as soon as possible (Annexin V- FITC can be imaged using FITC settings, while PI can be imaged using Cy@3 or Texas Red@ settings).

2. For adherent cells, the suggested protocol is as below:

1) Grow cells on coverslips or chamber slides.

2) Induce apoptosis in cells using the desired method. Prepare a negative control by incubating cells in the absence of inducing agent.

3) Wash cells with PBS twice, either preheated or at room temperature.

4) Prepare working solution: add 5 μ L Annexin V- FITC and 2 μ L PI to each 100 μ L 1×Annexin V Binding Buffer and mix gently.

5) Add appropriate amounts of working solution to cells and incubate at room temperature for 15 min in the dark (Incubation can be carried out on ice to slow down the apoptotic process, but the incubation time is extended to at least 30 min).

6) Wash cells with 1×Annexin V Binding Buffer twice.

Note: Do not use PBS to wash cells during this step.

7) Mount coverslips onto slides with a drop of 1×Annexin V Binding Buffer. For cells on chamber slides, add enough 1×Annexin V Binding Buffer to completely cover cells.

Note: Anti-fluorescence quenching agent can also be used.

8) Analyze cells by fluorescence microscopy using the appropriate filters as soon as possible.

Typical Data

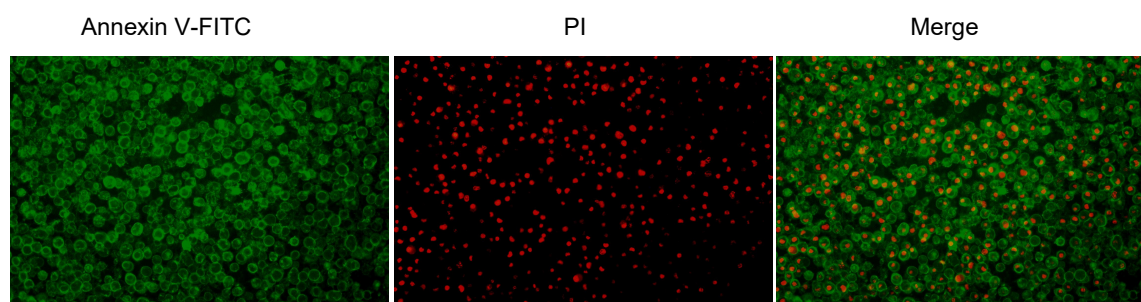


Figure 1. L929 cells were induced by this kit and were stained with Annexin V-FITC/PI Apoptosis Detection Kit to detect the apoptotic effect.

Recommended Products

Catalog No.	Product Name
KTA0001	Annexin V-AbFluor™ 405 Apoptosis Detection kit (Blue Fluorescence)
KTA0002	Annexin V-AbFluor™ 488/PI Apoptosis Detection kit
KTA0004	Annexin V-AbFluor™ 647 Apoptosis Detection kit (Red Fluorescence)
KTA0005	Annexin V-EGFP/PI Apoptosis Detection kit
KTA2010	One-step TUNEL Apoptosis Assay Kit (Green Fluorescence)
KTA2011	One-step TUNEL Apoptosis Assay Kit (Red Fluorescence)
KTA4001	Mitochondrial Membrane Potential Assay Kit (JC-1)

Disclaimer

The reagent is only used in the field of scientific research, not suitable for clinical diagnosis or other purposes.